Hashish:¹ Factors Influencing Double-Bond Stability in Cannabinoids¹

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It is shown that Bucourts methods for estimating torsion strain can be successfully applied to the THC ring system to give a surprisingly good quantitative estimate of the relative stability of various THC double bond isomers (Table II). trans-C1-Deoxy-THC's 2a and 2b are synthesized from the corresponding THC's by conversion to their phenyl ethers followed by cleavage with Na/NH_3 . The C₆-demethyl-THC's in the trans (3a and 3b) and cis (5a and 5b) series are synthesized from the pyrone (6) as shown in Scheme I.

A problem of long standing in the cannabinoid field has been the lack of knowledge of the factors which control the relative thermodynamic stabilities of the Δ^9 and Δ^8 double bond isomers in tetrahydrocannabinols (THC's).² A thorough understanding of these factors is desirable because the main physiologically active constituent of marijuana (Cannabis sativa L), Δ^9 -6a,10a-trans-THC (Table I, 1b), and its major metabolites in man and laboratory animals possess the thermodynamically less stable Δ^9 -unsaturation.

It is well-known that treatment of Δ^9 - or Δ^8 -6a,10atrans-THC (Table I; 1b and 1a) with acid results in an equilibrium mixture containing approximately 3% of the $\Delta^{\bar{9}}$ and 97% of the Δ^{8} isomer. The same isomer ratio was observed when the corresponding acetates were similarly treated. On the other hand, we have recently shown that in the cis-THC's (as their acetates)³ equilibration with acid gives 77% of the Δ^9 and 23% of the Δ^8 isomer (Table I; 4b and 4a).4

Previous attempts to rationalize these findings were based on steric arguments. In the preferred conformation of Δ^9 -6a,10a-trans-THC (Figure 1; derived from Westheimer calculations and nuclear Overhauser effects) substantial steric interactions exist between the C10 proton and the phenolic oxygen ($r_{\rm H_{10}-O_1} = 2.30$ Å), and the C10a proton and the C6 α -methyl group ($r_{\rm H_{10a}-C_{12}} = 2.89$ Å).⁵ It was suggested^{4,6} that these interactions are relieved by isomerization to the Δ^8 isomer, thus accounting for the preponderance of this isomer in the equilibrium mixture. Presumably these steric effects are absent in the cis isomers. To verify the validity of these arguments, we synthesized the C_1 -deoxy (2) and C_6 -didemethyl (3 and 5) THC's to minimize the nonbonded interactions. In light of the equilibrium data obtained for these cannabinoids (Table I), it is evident that the steric arguments are no longer tenable. Thus, acid treatment of Δ^8 - and Δ^9 trans-deoxy-THC's (2a and 2b) and Δ^{8} - and Δ^{9} -trans-didemethyl THC's (3a and 3b) does not significantly alter the equilibrium ratios when compared with their more hindered counterparts.

We have noted that the double-bond equilibria in the 6a,10a-cis- and trans-THC's are qualitatively similar to those in the corresponding octalins.⁴ It appears, then, that the methods pioneered by Barton and Corey and later expanded by Bucourt and Robinson⁷ to analyze conformational deformations and predict the relative stability of double-bond isomers in octalins, hexahydronaphthalenes, and other polycyclic molecules can be extended to the THC's. These considerations satisfactorily explain the results obtained to date in cannabinoids and are presented in this paper.

Discussion

As a result of work on triterpenoid ketones, Barton^{7b}

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Table I. Amounts of Various Cannabinoids under Equilibrium Conditions



postulated that conformational deformations brought about by the introduction of unsaturated substituents were

⁽¹⁾ Part 26. For part 25, see: Duffley, R. P.; Handrick, G. R.; Uliss, D. B.; Lambert, G.; Dalzell, H. C.; Razdan, R. K. Synthesis 1980, 733. Presented in part at the John C. Sheehan Symposium held at Hoboken, NJ, on Mar 20, 1980.

⁽²⁾ Reviews: (a) Razdan, R. K. Prog. Org. Chem. 1973, 8, 78-101; (b) Mechoulam, R., Ed. "Marijuana, Chemistry, Pharmacology, Metabolism and Clinical Effects"; Academic Press: New York, 1973; pp 2-100. (c) Mechoulam, R.; McCallum, N. K.; Burstein, S. Chem. Rev. 1976, 76, 75.

⁽³⁾ This equilibrium study was carried out on the acetates of cis-THC's since it is known that on acid treatment, cis-THC's undergo rearrangement to iso-THC's.

 ⁽⁴⁾ Uliss, D. B.; Razdan, R. K.; Dalzell, H. C.; Handrick, G. R. Tetrahedron Lett. 1975, 4369; Tetrahedron 1977, 33, 2055.
 (5) Archer, R. A.; Boyd, D. B.; Demarco, R. V.; Tyminski, I. J.; Al-

linger, N. L. J. Am. Chem. Soc. 1970, 92, 5200.

⁽⁶⁾ Gaoni, Y.; Mechoulam, R. J. Am. Chem. Soc. 1966, 88, 5673.

^{(7) (}a) Corey, E. J.; Sneen, R. A. J. Am. Chem. Soc. 1955, 77, 2505. (b) Barton, D. H. R.; Head, A. J.; May, P. J. J. Chem. Soc. 1957, 935. Bucourt, R. Top. Sterochem. 1974, 8, 159-224 and references therein. (d) Robinson, M. J. T.; Whalley, W. B. Tetrahedron 1963, 19, 2123.

Table II. Observed and Calculated Equilibrium Amounts^a

			obsd amt		calcd amt		
compd	$\theta, b \deg$	$\phi_{AO},^{b} \deg$	Δ9	Δ ⁸	Δ9	Δ^8	ΔE , kcal/mol
trans-THC (1)			3	97			-2.4
trans-1-deoxy-THC (2,			3	97			$^{-2.4}$
trans-6,6-didemethyl-THC (3)			3	97			-2.4
	110	45			6.0	94.0	-1.92
	113	45			3.1	96.9	-2.40
cis-THC (4)			77	23			0.84
cis-6,6-didemethyl-THC (5)			58	42			0.23
, ,		45			81.3	18.7	1.02
		46.4			77.1	22.9	0.85

 $^{a} \phi_{BO}$ taken as 45° and 61° for Δ ° and Δ ° type fusion, respectively. b Angles used in calculating equilibrium amounts, using eq 1 and 2.

Δ



Figure 1.

transmitted through the molecule by "flexing valency angles". This concept, known as conformational transmission, has since been interpreted in terms of the distortion of torsion (dihedral) angles⁸ at ring junctions from strain-free values. A semiquantitative method for estimating torsion strain has been developed by Bucourt^{7c} and successfully used to predict the relative stabilities of double-bond isomers in a variet of carbocyclic systems including octalins and hexahydronaphthalenes. Because we have shown that reducing selected intramolecular interactions in the THC's has little or no effect on the equilibrium ratio of the Δ^8 and Δ^9 double bond isomers, it may be that torsion strain about the 6a,10a fusion is the determining factor.

The use of torsion angles for estimating conformational energetics in the THC's is illustrated in Figure 2. The THC system can be considered as a fusion of a cyclohexene molecule and a benzopyran system as shown in Figure 2. The unfused torsion angle for the benzopyran system is approximated as 45° , the value for cyclohexene. This assumes that the substitution of carbon by oxygen in the benzopyran system has minimal effect on the torsion angle.

The amount of change in the torsion angles at the ring junction introduced in making the fusion is a measure of the torsional strain of the fused system. The geometry of fusion requires that for a trans fusion the sum of the absolute value of the two torsion angles at the junction equals a constant (approximately 110° when the substituents at a trans junction are hydrogen) while for a cis fusion the two torsion angles are equal. Thus, in the trans series the Δ^9 - and Δ^8 -THC analogues have torsion strains of 20° [110 - (45 + 45)] and 4° [110 - (45 + 61)], respectively. In the cis series the Δ^9 and Δ^8 analogues have torsion strains of 0° (45 - 45) and 16° (61 - 45), respectively. The above torsional considerations agree qualitatively with the observed stabilities of THC isomers. Namely, that trans-

 Δ^8 -THC is more stable than trans- Δ^9 -THC and cis- Δ^9 -THC is more stable than cis- Δ^8 -THC.

In addition to the above qualitative conclusions, we have found that very straightforward torsion angle energy considerations give a surprisingly good quantitative estimate of the relative stability of THC double-bond isomers. An estimate of the energy increase introded in a cyclohexane ring by a change $(\Delta\phi)$ in a torsion angle from its equilibrium value has been given by Bucourt as $(\Delta\phi)^2/100$ for an opening and $2/3(\Delta\phi)^2/100$ for a closing (energy in kilocalories and $\Delta\phi$ in degrees) of the torsion angle. When these functions are summed for the two rings involved in a fusion and then minimized, eq 1 and 2 are obtained, $\Delta H_{Yel} =$

$$\frac{1}{100} \left(\frac{f_{A}f_{B}}{f_{A} + f_{B}} \right) \left[\theta - (\phi_{AO} + \phi_{BO}) \right]^{2} \text{ for trans fusion (1)} \\
H_{Kcal} = \frac{1}{100} \left(\frac{f_{A}f_{B}}{f_{A} + f_{B}} \right) (\phi_{BO} - \phi_{AO})^{2} \text{ for cis fusion (2)}$$

representing the torsion strain introduced in trans and cis ring fusions, respectively. (ϕ_{AO} and ϕ_{BO} are the equilibrium torsion angles at the junction in degrees for rings A and B before fusion; f_A (and f_B) equals 1 if fusion requires an angle opening in that ring and 2/3 if a closing; θ is approximately 110°.)

The torsion strain and calculated equilibrium values obtained by using these equations are compared with experimental values in Table II. Also shown in Table II are values of θ and ϕ_{AO} that would lead to agreement between calculated and observed equilibrium constants for *trans*-THC's and *cis*-THS's. These values (ϕ of 113° and ϕ_{AO} of 46.4°) are not at all unreasonable as starting parameters.

The agreement of calculated and experimental equilibrium values is remarkable considering the simplicity of the energy calculations. Implicit in these calculations is the assumption that the relative energy of the THC doublebond isomers can be approximated by torsion strain at the ring junction, using a torsion energy function for cyclohexane. It is further assumed that no change in entropy occurs during the isomerizations. While this assumption may be reasonable for trans-THC's, being quite rigid with a single conformer highly favored, it may not be so for cis-THC's, which appear more flexible. We speculate that entropy effects may, in fact, be responsible for the different equilibrium values found for cis-THC and cis-6,6-didemethyl-THC. The lack of the gem-dimethyl groups in the latter may allow the population of additional conformations.

The results reported here are a further example of the value of simple ring-junction torsion angle considerations in the evaluation of energetics of multiring systems.

⁽⁸⁾ Defined as the angle subtended by the planes of two vicinal bonds when viewed along the connecting bond in the Newman projection. An ortho ring fusion contains two such angles. See: Klyne, W.; Prelog, V. Experientia 1960, 16, 521.

trans fusion



Synthesis

The C_1 -deoxy THC's in the trans series (2a and 2b) were synthesized from the corresponding THC's by conversion to their phenyl ethers followed by cleavage of the phenyl ether with sodium and liquid ammonia. In the Δ^8 series, compound 2a was formed in good yield and was identical with the compound prepared by Kraatz and Korte⁹ by a different route. However, in the Δ^9 series, the Na/NH₃ reaction gave a complex mixture from which 2b was isolated in poor yield. It was only $\sim 60\%$ pure by GLC and could not be purified further despite the repeated use of high-pressure liquid chromatography (LC). The structure was confirmed by comparison of its NMR spectra with those of 2a. The vinylic proton in 2b was observed at δ 5.93 compared to δ 5.38 in 2a. The major product formed during this reduction has been tentatively identified as 1-deoxyhexahydrocannabinol (see Experimental Section).

The C₆-didemethyl THC's in the trans (**3a** and **3b**) and cis (**5a** and **5b**) series were synthesized from the known pyrone 6^{10} following the sequence as shown in Scheme I.

The pyrone 6 with LiAlH₄ in ether formed the triol 7 which on treatment with thionyl chloride/triethylamine in CH₂Cl₂ followed by hydrolysis of the ketal with 6 N HCl furnished 8. Reduction with Li/liquid NH₃ converted 8 into a mixture of cis and trans ketones 9b and 9a respectively, which were separated as their acetates 10b and 10a by using high-pressure LC. Pure 9a and 9b were obtained by saponification of the corresponding acetates 10a and 10b. Grignard addition to 10a and 10b gave 11a and 11b, respectively, which were dehydrated with thionyl chloride in pyridine to yield a mixture of Δ^8 - and Δ^9 -THC's. Thus 11a gave a mixture of 3a and 3b whereas 11b gave a mixture of 5a and 5b. The trans and cis ring junctures of 3 and 5 were assigned on the basis of the position of the olefinic protons at C10 and C8 in the NMR.^{4,10} In the trans series (3b and 3a) they were observed at δ 6.37 and 5.38, respectively, compared to δ 5.53 and 5.30 in the cis series (5b and 5a).

It is interesting to note that cis-didemethyl-THC's (5), unlike cis-THC's, do not undergo rearrangement to iso-

⁽⁹⁾ Kraatz, U.; Korte, F.; Tetrahedron Lett. 1976, 1977.

⁽¹⁰⁾ Fahrenholtz, K. E.; Lurie, M.; Kierstead, R. W. J. Am. Chem. Soc. 1967, 89, 5938.

THC's in the presence of acids. This is not surprising because the necessary cleavage of the C–O bond requires formation of the unfavorable primary carbonium ion in the former as opposed to a tertiary carbonium ion in the latter.



Experimental Section

All compounds are (\pm) racemic mixtures. Melting points were determined in a Thomas-Hoover melting-point apparatus and are uncorrected. The IR spectra were recorded on a Perkin-Elmer Model 700 instrument and the NMR spectra were measured on a Varian T-60 spectrometer. The high-pressure liquid chromatographic separations were made with a Waters Associates ALC-202 chromatograph equipped with a Model 6000 solvent delivery system. Preparative separations were made on a 7 ft \times 0.375 in. i.d. column packed with Porasil C. Analytical separations were made on a 1 ft \times 0.25 in. o.d. column packed with μ Porasil. The analyses by gas chromatography were made on a Varian Aerograph Model 1440, equipped with a 6 ft \times 0.125 in. i.d. stainless-steel column packed with 2% OV-17 on 100/200-mesh Supelcoport and a flame-ionization detector. Compounds 4a and 4b were prepared according to the procedure described by us earlier.4

Preparation of 6a,10a-*trans*-3-Pentyl-6a,7,10,10a-tetrahydro-6,6,9-trimethyl-6*H*-dibenzo[*b*,*d*]pyran (2a). Refluxing 1a (0.51 g, 1.62 mmol) with 1 mL of bromobenzene in 12 mL of pyridine containing 117 mg of copper powder and 268 mg of anhydrous potassium carbonate, according to a procedure of Sawa,^{11a} gave the phenyl ether of 1a in 65% yield. The analytical sample was obtained as a colorless oil after passage through a short Florisil column with petroleum ether; NMR (CCL₄) & 0.87 (t, 3, 5'-CH₃), 1.07, 1.33 (2 s, 6, 6-C(CH₃)₂), 1.60 (s, 3, 9C-CH₃), 2.42 (t, 2, 1'-CH₂), 2.97 (dd, 1, J = 4, 17 Hz, 10-CH_a), 5.30 (br, 1, 8-CH), 6.18, 6.37 (2 d, J = 2 Hz, 2-CH, 4-CH), 6.8–7.4 (m, 5, OC₆H₆). Anal. Calcd for C₂₇H₃₄O₂: C, 83.03; H, 8.77. Found: C, 83.11; H, 9.04.

Cleavage of the phenyl ether (0.50 g, 1.28 mmol) with Na/ NH₃^{11b} gave **2a** in 63% yield as a colorless oil which was pure by GLC (retention time 2.75 min; column temperature 240 °C); NMR (CCl₄) δ 0.88 (t, 3, 5'-CH₃), 1.13, 1.37 (2 s, 6, 6-C(CH₃)₂), 1.70 (s, 3, 9C-CH₃), 2.47 (t, 2, 1'-CH₂), 2.63 (d, J = 4 Hz, 10a-CH), 5.38 (br s, 1, 8-CH), 6.50 (d, $J_{2,4} = 1$ Hz, 1, 4-CH), 6.60 (dd, $J_{1,2} = 8$, $J_{2,4} = 1$ Hz, 1, 2-CH), 6.97 (d, $J_{1,2} = 8$ Hz, 1, 1-CH). Anal. Calcd for C₂₁H₃₀O: C, 84.50; H, 10.13. Found: C, 84.24; H, 10.27.

These data agree with those of 2a prepared by Kraatz and Korte⁹ by a reductive cleavage of a phosphate ester of $(-)-\Delta^8$ -THC by Li/NH₃.

Preparation of 6a,10a-trans-3-Pentyl-6a,7,8,10a-tetrahydro-6,6,9-trimethyl-6*H*-dibenzo[*b*,*d*]pyran (2b). The phenyl ether of 1b was prepared in 70% yield in the manner described for the Δ^6 isomer. Chromatography on Florisil using 1:1 benzene-petroleum ether as eluant gave the analytical sample; NMR (CCl₄) δ 0.87 (t, 5'-CH₃), 1.03, 1.37 (2 s, 6, 6-C(CH₃)₂), 2,43 (t, 2, 1'-CH₃), 3.00 (br d, J = 9 Hz, 1, 10a-CH), 6.2 (br, 1, 10-CH), 6.27, 6.40 (2 d, J = 2 Hz, 2-CH, 4-CH), 6.7-7.4 (m, 5, C₆H₈). Anal. Calcd for C₂₇H₃₄O₂·H₂O: C, 79.37; H, 8.88. Found: C, 79.39; H, 8.71.

Cleavage of the ether as described above and separation of products by preparative thick-layer chromatography gave 2b (9%, 60% pure by GLC) and two other compounds tentatively identified as 1-deoxycannabinol (5%) and 1-deoxyhexahydrocannabinol (48%). Compound 2b could not be purified further despite repeated high-pressure liquid chromatography: NMR (CCl₄) δ 0.88 (t, 3, 5'-CH₃), 1.13, 1.37 (2 s, 6, 6-C(CH₃)₂), 1.70 (s, 3, 9C-CH₃), 2.47 (t, 2, 1'-CH₂), 3.27 (br d, 1, J = 24 Hz, 10a-CH), 5.93 (br s, 1, 10-CH), 6.53 (d, $J_{2,4} = 1$ Hz, 1, 4-CH), 6.58 (dd, $J_{1,2} = 8$, $J_{2,4} = 1$ Hz, 1, 2-CH), 7.10 (d, $J_{1,2} = 8$ Hz, 1, 1-CH); GLC retention time 2.99 min (column temperature 240 °C).

Preparation of 7-(5'-Pentyl-2'-resorcinyl)-1,4-dioxaspiro-[4,5]dec-7-ene-8-methanol (7). A solution of 6¹⁰ (6.88 g, 0.02 mol) in 70 mL of benzene and 10 mL of ether was added dropwise to a slurry of lithium aluminum hydride (3.20 g, 0.08 mol) in 250 mL of ether at room temperature. The mixture was brought to reflux for 45 min and cooled to 0 °C, and excess hydride was destroyed by addition of 10 mL of ethyl acetate in 6 mL of ether followed by 40 mL of saturated aqueous ammonium chloride. The organic layer was decanted and the residue was dissolved by addition of ammonium chloride and water. The aqueous layer was washed with ether, and the combined organic layer was washed with water and saturated sodium chloride solution, dried, and concentrated. The residue crystallized when a solution in ether was mixed with excess petroleum ether. The yield of triol 7 was 6.23 g (89%). The analytical sample, mp 111.5-112 °C, was obtained by recrystallization from ether: IR (CDCl₃) ν 1040, 1440, 1570, 1625, 3450 (OH) cm⁻¹; NMR (CDCl₃) & 0.87 (t, 3, 5"-CH₃), 1.38 (m, 6), 1.92 (m, 2, 10-CH₂), 2.2–2.8 (m, 6, 1"-CH₂, 6-CH₂, 9-CH₂), 3.90 (s, 2, α -CH₂), 4.02 (s, 4, 2-CH₂, 3-CH₂), 6.35 (s, 2, 4'-CH, 6'-CH), exchangeable OH at δ 3.1, 6.3. Anal. Calcd for C₂₀H₂₈O₅: C, 68.94; H, 8.10. Found: C, 68.89; H, 8.09.

Preparation of 9-Oxo-3-pentyl-6a,7,8,9-tetrahydro-6Hdibenzo[b,d]pyran-1-ol (8). A solution of thionyl chloride (1.90 mL, 0.026 mol) in 23 mL of methylene chloride was added over 1.5 h to a solution of 7 (6.00 g, 0.017 mol) and triethylamine (24 mL, 0.17 mol) in 90 mL of methylene chloride cooled to -20 °C under nitrogen. The reaction mixture was allowed to warm to 5 °C and then poured onto 200 mL of ice and water. The organic layer was washed with water, dilute HCl, dilute sodium bicarbonate, and water. Drying over Na2SO4 and removal of solvent gave an oil which solidified on standing. A solution of the solid in dioxane (50 mL) was mixed with 20 mL of 6 N HCl and the two-phase mixture was heated to 80 °C for 75 min. The reaction mixture was then diluted with an equal volume of water and the dioxane removed in a rotary evaporator. Crude ketone 8 (4.62 g) was collected by filtration and purified by extraction from tar with hot CCl₄. Addition of ether to the CCl₄ extract precipitated 8 in 56% yield: mp 172-175 °C; IR (CDCl₃) v 1610 (C=0, conjugated ketone), 3150 (OH, strongly hydrogen bonded) cm⁻¹; NMR (CDCl₃) & 0.88 (t, 3, 5'-CH₃), 3.73, 4.30 (m, 2 H, 6-CH₂), 6.32, 6.60 $(2 d, 2, 2-CH, 4-CH), 7.95 (d, 1, J = 2 Hz, 10-CH).^{10}$ Anal. Calcd for C₁₈H₂₂O₃: C, 75.49; H, 7.74. Found: C, 75.17; H, 7.48.

Preparation of 6a,10a-trans- and -cis-6a,7,8,9,10,10a-Hexahydro-9-oxo-3-pentyl-6H-dibenzo[b,d]pyran-1-ol Acetates (10a and 10b). Lithium wire was added to 200 mL of liquid ammonia until a permanent blue color was obtained. Compound 8 (2.70 g, 0.0094 mol) in 30 mL of tetrahydrofuran was added over 1 h. During the addition a further 0.74 g (0.01 mol) of lithium was added to maintain the blue color. Excess ammonium chloride was added and the ammonia allowed to evaporate. Water and ether were added, the organic layer was washed with water, dilute HCl, dilute NaHCO₃ solution, and brine and dried over Na₂SO₄ and solvent was removed. The residue was 3.10 g of a solid, which was passed through a short column of Florisil with methylene chloride and crystallized from ether-petroleum ether to give a mixture of 9a and 9b (2.0 g, 76%).

A mixture of the corresponding acetates (10a and 10b) was prepared in 75-85% yield by reaction with acetic anhydride in pyridine. Separation was effected by high-pressure liquid chromatography using a Porasil column and 1:2 ether-isooctane as eluant; k' was 2.00 and 2.47 for 10a and 10a, respectively. Analytical samples were obtained from the concentrated eluants by trituration with petroleum ether (10a, mp 101-103 °C; 10b, mp 59-61 °C). 10a: NMR (CDCl₃) δ 0.87 (t, 3, 5'-CH₃), 2.32 (s, 3, CO₂CH₃), 3.70 (dd, 1, J = 10, 10 Hz, 6α -CH), 4.25 (dd, 1, J =10, 4 Hz, 6β -CH), 6.43, 6.62 (2 d, 2, J = 2 Hz, C2-H, C4-H). The NMR spectrum of 10b was very similar to that of 10a except that 6α -CH and 6β -CH appeared as a multiplet at δ 4.20. Anal. Calcd for C₂₀H₂₆O₄: C, 72.70; H, 7.93. Found for 10a: C, 72.72; H. 7.95. Found for 10b: C, 72.72; H, 7.94.

Preparation of 6a,10a-*trans*- and -*cis*-6a,7,8,9,10,10a-Hexahydro-9-oxo-3-pentyl-6*H*-dibenzo[*b*,*d*]pyran-1-ols (9a

^{(11) (}a) Sawa, Y. K.; Tsuji, N.; Maeda, S. Tetrahedron 1964, 20, 2255;
(b) Ibid. 1961, 15, 144, 154.

Table III. Equilibration Studies with Tetrahydrocannabinols^c

with fellanyarocannabinois									
compd	time, min	% Δ ⁸	% Δ°						
1a	0	100							
	4-1350	97	3						
1a acetate	0	100							
	4-1350	97	3						
1b	0	0	100						
	4	93	7						
	30	97	3						
1b acetate	0	0	97						
	4	32	68						
	10	78	22						
	30	97	3						
2a	0-1080	100	0						
	0	0	100 <i>ª</i>						
	5	97	3						
	10	97	3						
	35	97	3						
3	0	83	17						
	20	97	3						
4a acetate	0	93	7						
	30	87	13						
	60	80	20						
	120	77	23						
	222 ^b	80	20						
	300	28	72						
	336	23	77						
4b acetate	0	0	100						
	30	13	87						
	60	23	77						
5	0	56	44						
	20	53	47						
	180	45	55						
	390	42	58						

^a Material could not be purified >60% (GLC). It did not contain any Δ^{s} isomer. ^b More *p*-TSA added at 270 min. ^c Refluxing dilute solutions in benzene with *p*-TSA.

and 9b). Pure 9a and 9b were obtained by saponification of the corresponding acetates (10a and 10b) with 5% KOH in methanol at room temperature for 15 min. Analytical samples were recrystallized from ether-petroleum ether (9a, mp 173 °C; 9b, mp 161 °C); 9a: IR (CDCl₃) ν 1685 (C=O), 3270 (OH) cm⁻¹. 9b: 1700 (C=O), 3320 (OH) cm⁻¹. Anal. Calcd for C₁₈H₂₄O₃: C, 74.97; H, 8.39. Found for 9a: C, 74.77; H, 8.45. Found for 9b: C, 74.83; H, 8.40.

Preparation of 6a,10a-trans- and -cis-6a,7,8,9,10,10a-Hexahydro-9-methyl-3-pentyl-6H-dibenzo[b,d]pyran-1,9diols (11a and 11b). A solution of 10a (0.55 g, 0.0016 mol) in 6 mL of ether and 2 mL of benzene was added over 10 min under nitrogen to a solution of methylmagnesium iodide, prepared from 0.40 g (0.016 mol) of magnesium turnings and 1.2 mL (2.36 g, 0.0166 mol) of iodomethane, in 9 mL of ether. After 60 min the cooled reaction mixture was quenched by successive addition of ethyl acetate and methanol in ether and saturated ammonium chloride. The organic layer was washed with dilute sodium chloride, dried, and concentrated to give 11a (88%) as a resin (mixture of isomers at C₉ which was not separated). The analytical sample was obtained by addition of petroleum ether to an ethereal solution off 11a: mp 141-142 °C; NMR (CDCl₃) δ 0.87 (t, 3, 5'-CH₃), 1.37 (s, 3, 9-CH₃), 2.42 (t, J = 6 Hz, 2), 3.2-4.2 (m, 3), 6.10, 6.25 (2 d, J = 2 Hz, 2, 2-CH, 4-CH), 2.01, 6.82 (br, 2, exchangeable OH); IR (CDCl₃) ν 3330 (OH) cm⁻¹. Anal. Calcd for C₁₉H₂₈O₃: C, 74.96; H, 9.27. Found: C, 75.05; H, 9.29.

In a similar fashion, 11b was obtained from 10b in 97% yield (mp 131-133 °C): NMR (CDCl₃) δ 0.87 (t, 3, 5'-CH₃), 1.38 (s, 3, 9-CH₃), 4.0 (d, J = 4 Hz, 1), 4.25 (t, J = 10 Hz, 1), 4.70 (m, 1), 6.15, 6.28 (2 d, J = 2 Hz, 2, 2-CH, 4-CH), 1.8, 5.9 (br, 2, exchangeable OH); IR (CDCl₃) ν 3350 (OH) cm⁻¹. Anal. Calcd for C₁₉H₂₈O₃: C, 74.96; H, 9.27. Found: C, 74.85; H, 9.29.

Preparation of 6a,10a-trans-9-Methyl-3-pentyl-6a,7,10,10aand -6a,7,8,10a-tetrahydro-6H-dibenzo[b,d]pyran-1-ols (3a and 3b). To an ice-cold solution of 11a (0.100 g, 0.320 mol) in a mixture of 1 mL of methylene chloride and 2 mL of pyridine was added a solution of thionyl chloride (0.16 g, 0.001 37 mol) in 1 mL of methylene chloride. After 1 h, 10 mL of ether and 6 mL of water were added. The organic layer was washed with water, dilute HCl, dilute NaHCO₃ solution, water, and saturated sodium chloride solution. Drying and removal of solvent left 91 mg of a compound tentatively identified as a cyclic sulfite ester. This material was hydrolyzed with 5% KOH in methanol and enough ether was added for homogeneity. After 1 h at room temperature ether was added. The ether layer was washed with water, dilute HCl, dilute NaHCO₃, and brine. Drying and evaporation of solvent left 92 mg of resin which was purified by column chromatography (Florisil, 2% ether-petroleum ether). A fraction was obtained which consisted of 83% 3a and 17% 3b (GLC); NMR (CCl₄) δ 0.88 (t, 3, 5'-CH₃), 1.68 (s, 3, 9-CH₃), 5.38 (br, olefinic, 8-CH), 5.98, 6.18 (2 d, J = 2 Hz, 2, 2-CH, 4-CH), 6.37 (br, olefinic, 10-CH), 5.03 (s, 1, exchangeable OH). Anal. Calcd for C19H28O2: C, 79.67; H, 9.15. Found: C, 79.47; H. 9.25.

By a similar series of reactions, 100 mg of 11b was converted to a mixture of 53% **5a** and 47% **5b** (GLC) in 62% yield; NMR (CCl₄) δ 0.90 (t, 3, 5'-CH₃), 1.63 (s, 3, 9-CH₃), 5.30 (br, olefinic, 8-CH), 5.53 (br, olefinic 10-CH), 6.00, 6.17 (2 d, 2, 2-CH, 4-CH), 4.83 (s, 1, exchangeable OH). Anal. Calcd for C₁₉H₂₆O₂-0.5H₂O: C, 77.25; H, 9.21. Found: C, 77.58; H, 8.98.

Equilibration of Tetrahydrocannabinols. The general procedure was to reflux under nitrogen a stirred solution of the THC's (approximately 0.01 mol) in dry benzene containing *p*-toluenesulfonic acid monohydrate as the acid catalyst. Samples for GLC analysis, taken at regular intervals, were treated with solid Na₂CO₃·H₂O before injection. The reaction was stopped when it appeared to reach equilibrium, and solid Na₂CO₃·H₂O was added to neutralize the acid catalyst. The products were identified by comparison with authentic specimens from other sources, both unsilylated and silylated analyses being required in the GLC because of overlapping retention times.

Treatment of a mixture containing 53% 5a and 47% 5b (GLC) with excess *p*-TSA in refluxing benzene for 6.5 h showed that no other product had formed during the reaction (NMR, GLC) and only the ratio of 5a and 5b had changed to 42% and 58%, respectively.

Equilibration studies showing composition vs. time for various THC's are shown in Table III.

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Registry No. (±)-1a, 6087-61-2; (±)-1a acetate, 76248-17-4; (±)-1a phenyl ether, 76207-25-5; (±)-1b, 3556-79-4; (±)-1b acetate, 76248-18-5; (±)-1b phenyl ether, 76207-26-6; (±)-2a, 76248-19-6; (±)-2a, 76207-27-7; (±)-3a, 76207-28-8; (±)-3b, 76207-29-9; (±)-4a, 6216-87-1; (±)-4a acetate, 58769-87-2; (±)-4b, 6087-73-6; (±)-4b acetate, 23050-53-5; (±)-5a, 76207-30-2; (±)-5b, 76207-31-3; (±)-6, 6469-57-4; (±)-7, 76207-32-4; (±)-8, 76207-33-5; (±)-9a, 76207-34-6; (±)-9b, 76207-35-7; (±)-10a, 76207-36-8; (±)-10b, 76207-37-9; 11a, 76207-38-0; 1-deoxytexahydrocannabinol, 76207-39-1; 1-deoxycannabinol, 76207-40-4; methyl iodide, 74-88-4.